IMMUNOLOGICAL TECHNIQUES IN CLINICAL DIAGNOSIS

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ANTIGEN PRODUCTION

- Method of choice depends upon organism, location of antigens in cell and type of antigen
- Cell fractionation
- Separation of desired antigen
- Purification
- Storage in small amounts
- Immunization of test animal

ANTIBODY PRODUCTION

Desired quantity of antigen is injected into test animal at multiple points with Freund's complete Adjuvant



Booster doses are given after testing antibody production on each dose of immunization

Last doze is given with Freund's Incomplete Adjuvant

Animal is bled aseptically by heart puncture

Antibodies are separated by centrifugation and stored in small quantities at -20°C



Antigen-Antibody-Ratio determines visible insoluble precipitate



precipitate

Basic Principle of Dilution



Precipitation Reaction



Antigens (soluble)

Cone of equivalence: Similar Structure Struc

Antibodies



Agglutination Reaction



Ochterlony Immuno-double Diffusion



DOUBLE IMMUNODIFFUSION





Principle of Electrophoresis



Immunoelectrophoresis



It combines electrophoresis separation, diffusion and precipitation of protein antigens

Rocket Immunoelectrophoresis

Crossed Immunoelectrophoresis

Secondary Antibodies Production

- Enhance sensitivity of immuno-reactions
- When antibodies of one animal are injected into another/second animal, they act as antigens in second animal
- Antibodies produced in second animal are called "secondary antibodies"
- Purified secondary antibodies are labeled with horse radish peroxidase enzyme / gold particles / ferritin molecules / fluorescent dyes

Immunofluorescence

Fluorescent Antibody Staining Giardia (left) Cryptosporidium (right)

Immunoblotting

- Proteins are separated on SDS-PAGE
- Immobilized on Nitrocellulose membrane
- Membrane is cut in two pieces-one containing marker + proteins; another only protein bands
- Additional sites blocked with 2% skimmed milk
- Second piece cut into 5 mm wide stripes,
- Reaction allowed with primary Ab, washed with PBS + Tween-20 followed by reaction with secondary Ab, chromogen substrate added and reaction developed with H_2O_2 .
- Bands observed, molecular wt. determined

Major Steps in Immunoblotting

Typical Western Blot of HIV patients

Immunoblot-when proteins are not dominant antigens

PLATE 71 : Immunoblot against mycelial antigens of Trichopyton mentagrophytes.

P = Pre-immune serum ; U = Unabsorbed serum ; Ah = Absorption of serum with heat-killed homologous mycelium ; Af = Absorption of serum with fresh homologous mycelium ; M = Marker ; An = Antigen ; Tm mix = Mixture of all sera raised against T. mentagrophytes ; Tr mix = Mixture of all sera raised against T. rubrum ; Mc mix = Mixture of all sera raised against M. canis ; Mg mix = Mixture of all sera raised against M. gypseum.

ENZYME LINKED IMMUNOSORBANT ASSAY

(ELISA)

Immobilised Antigen	Primary Antibody	Conjugated Secondary Antibody	Substrate Addition Signal Detection & Quantification	
0) 0) 0)		Wash	Wash	

RIA and DOT BLOT are the variations of ELISA and Immunoblot respectively.

IMMUNO-ELECTRON MICROSCOPY

- **PRE-EMBEDDING:** Cells treated with primary antibodies, washed, treated with secondary antibodies labeled with gold particles or ferritin molecules
- **POST-EMBEDDING:** Ultra thin sections allowed to react with primary antibodies followed by washing, reacted with secondary antibodies labeled with gold particles or ferritin molecules
- Normal Electron Microscopy followed

PLATE 83 : T. mentagrophytes, Untreated Control, I. Absorbed, Rabbit-Anti-T.m. II. Anti-Rabbit-Ig-F , Moderate Ig-binding , Cells are healthy (0.M. 10000 x, F.M. 26500 x, 23696).

PLATE 84 : T. mentagrophytes, Untreated Control, I. Absorbed, Rabbit-Anti-T.m. II. Anti-Rabbit-Ig-F , Moderate Ig-binding , Cells are healthy (0.M. 30000 x, F.M. 80800 x, 23692).

PLATE 81 : T. mentagrophytes, Untreated Control, I. Unabsorbed, Rabbit-Anti-T.m. II. Anti-Rabbit-Ig-F,Strong Ig-binding, Cell is healthy and normal (0.M. 30000 x, F.M. 87000 x, 23371).

THANKING YOU

